The role of presepsin in the diagnosis of chronic obstructive pulmonary disease acute exacerbation with pneumonia

Ali Halıcı¹, İzzettin Hür¹, Kerim Abatay¹, Esra Çetin², Filiz Halıcı³ & Seda Özkan*,⁴

¹Department of Emergency Medicine, University of Health Sciences, Diskapi Yildirim Beyazit Training & Research Hospital, 06145, Ankara, Turkey

²Department of Biochemistry, University of Health Sciences, Diskapi Yildirim Beyazit Training & Research Hospital, 06145, Ankara, Turkey

³Department of Family Medicine, University of Health Sciences, Diskapi Yildirim Beyazit Training & Research Hospital, 06145, Ankara, Turkey

⁴Department of Emergency Medicine, University of Istanbul-Cerrahpasa, Faculty of Medical, 34098, Istanbul, Turkey *Author for correspondence: Tel.: +90 212 414 3434; sedacil@gmail.com

Aim: In this study, we aimed to investigate the role of presepsin in detecting concomitant pneumonia in patients presenting with acute exacerbation of chronic obstructive pulmonary disease (COPD) in the emergency department. **Patients & methods:** Three groups were formed in the study. Group 1: patients diagnosed with acute exacerbation of COPD; group 2: patients with acute exacerbation of COPD + pneumonia; group 3: healthy individuals. **Results:** Presepsin levels of the patients in group 2 were significantly higher than those of group 1 and group 3 (p < 0.05). There was a statistically significant difference in erythrocyte sedimentation rate, CRP, procalcitonin and presepsin values between two patient groups (p < 0.05). **Conclusion:** Presepsin can be used to diagnose pneumonia in patients with acute exacerbation of COPD admitted to the emergency department.

First draft submitted: 27 April 2019; Accepted for publication: 11 October 2019; Published online: 8 November 2019

Keywords: acute exacerbation • C-reactive protein • COPD • diagnosis • erythrocyte sedimentation rate • pneumonia • presepsin • procalcitonin

An exacerbation of chronic obstructive pulmonary disease (COPD) is defined as an acute worsening of respiratory symptoms that results in additional therapy [1,2]. In acute exacerbation of COPD, complicating events occur, such as an increase in airway inflammation and mucus production and an increase in aeration. These changes lead to an increase in dyspnea, a change in sputum volume and color, an increase in purulence, an exacerbation of cough and a high fever and/or impaired mental function [2]. Acute exacerbations of approximately one to four are observed in COPD patients every year [3]. Studies have shown that as the severity of COPD increases, the frequency of attacks increases. Infection and environmental factors play a role in the formation of acute exacerbations [4]. Bacterial and viral infections have an important role in the development of attacks [3]. However, 30% of the patients do not have a real reason to explain the exacerbation [4].

COPD is a risk factor for community-acquired pneumonia, and pneumonia is the most common cause of acute exacerbation of COPD [4]. Coexistence of COPD and pneumonia increases hospitalization and mortality rates [5]. Therefore, the Global Initiative for Chronic Obstructive Lung Disease (GOLD) guideline recommends initiation of antibiotics in the presence of purulent sputum in patients presenting with acute exacerbation of COPD [2].

In recent years, studies on the role of some systemic biomarkers in detecting etiology, prognosis and the risk of mortality in acute exacerbations of COPD are outstanding [6–9]. Procalcitonin and CRP are among these markers. Procalcitonin, which is accepted as a marker of bacterial infection, has been shown to correlate with the etiology and prognosis of pneumonia [6]. Procalcitonin is considered a guide in determining treatment with antibiotics because it can distinguish bacterial and viral infections [10]. CRP is an acute phase reactant that increases in cases of inflammation and infection [11]. CRP cannot differentiate bacterial infections from viral and other inflammatory



Biomarkers



conditions, but it is used to determine the prognosis of these conditions. CRP is much less affected in other conditions than other acute phase reactants, especially erythrocyte sedimentation rate (ESR) [11].

CD-14 is a protein of approximately 55 kDa that is present in the membranes of mononuclear cells, and it serves as a high-affinity receptor to lipopolysaccharides (LPS) and the LPS/LPS-binding protein complex (LPBP). The LPS-LPBP-CD14 complex is separated from the cell membrane and released into the circulation and forms sCD14. Another sCD14 produced by plasma protease activity is called sCD14-subtype (presepsin). Presepsin is a polypeptide with a weight of 13 kDa at the terminal end of CD14. Presepsin has no LPS-binding properties, so it cannot be captured by anti-CD14 antibodies; thus, this free molecule can be used as a biomarker. Numerous studies have shown that presepsin levels are increased in the early stages of bacteremia and sepsis, and presepsin is a biomarker that has been nominated for routine use [12–15].

Acute exacerbation of COPD is often accompanied by pneumonia, which changes the prognosis and treatment of the disease. In patients with acute exacerbation of COPD presenting to the emergency department, the diagnosis of concomitant pneumonia may be difficult.

In this study, we aimed to investigate the role of presepsin in detecting concomitant pneumonia in patients presenting with acute exacerbation of COPD to the emergency department.

Patients & methods

This observational prospective study was carried out in line with research regulations, including the approval of the Ethics Committee of our institute dated 19 March 2018 and numbered 47/10 and according to the principles of the 'World Medical Association Helsinki Declaration.' It was supported by scientific studies support board of our institute numbered 69/2.

The study included patients with acute exacerbation of COPD above 18 years of age. Patients under 18 years of age, those with malignancy, pulmonary embolism, pulmonary edema, bronchiectasis, other chronic lung disease, nonpneumonia infection and patients with sepsis were excluded from the study.

Three groups were formed in the study:

- Group 1: patients diagnosed with acute exacerbation of COPD (n = 65);
- Group 2: patients with acute exacerbation of COPD and pneumonia (n = 61);
- Group 3: healthy individuals (control; n = 44).

The healthy group was selected from volunteer patients' relatives and emergency department staff who did not have any disease. Patients who met the following criteria were diagnosed with acute exacerbation of COPD: exacerbated cough and dyspnea, increased sputum volume and increased sputum purulence.

In patients with acute exacerbation of COPD, new pulmonary infiltration in chest x-ray and/or computerized tomography (CT) was included in the COPD acute attack and pneumonia group. In addition to acute exacerbation symptoms of COPD, patients with new pulmonary infiltrations and signs of infection were included in group 2. Chest x-ray and/or CT was performed in all patients to detect pulmonary infiltration. Patients without a definite diagnosis of pneumonia and doubtful infiltrates were excluded from the study. Patients with pulmonary tuberculosis, pulmonary oedema, pulmonary thromboembolism, non-pneumonia infection were excluded. Patients using antibiotics were excluded from the study. All participants were informed about the study.

The patient's symptoms, vital signs, co-morbidities, demographic characteristics, symptoms and physical examination findings, laboratory results, imaging findings and treatments were recorded in the study form. There was no intervention in the follow-up and treatment of the patients. In addition to the routine blood tests performed for the patients, 5 ml of blood was taken from the patients to determine the serum presepsin level. Blood samples of the patients were collected at the time of admission to the emergency department before treatment was started. Blood samples were taken from healthy subjects only for presepsin levels. Blood samples taken from the patients were centrifuged at 3000 r.p.m. for 15 min, and the serum was pipetted into Eppendorf tubes and stored at -80°C until they were assayed. Serum presepsin levels were measured by ELISA, MBS766136, 0.1156–10 ng/ml in the biochemistry laboratory using a kit with a sensitivity of less than 0.094 ng/ml.

Other inflammatory markers (ESR, procalcitonin, CRP) were studied in the hospital laboratory with routine blood samples. Samples were tested for ESR (range: 0–20 mm/h) using a brand test 1 model device (Alifax S.p.A, Padova, Italy) with the Westergren method. CRP (range: 0–0.5 mg/dl) was studied in a Beckman Coulter Image 800 device (Beckman Coulter, CA, USA) with the nephelometric method. Procalcitonin was studied in an AQT90FLEX

analyzer device (Radiometer Medical Aps, Copenhagen, Denmark) with the time-resolved Fluorescence method (range: $<0.05 \mu g/l$).

Sputum samples of the patients were taken during the follow-up in the emergency department. Samples of sputum and tracheal aspirate were studied in our microbiology laboratory. Because of the intensity of the emergency room, only patients with acute exacerbation of COPD and pneumonia could be sampled for sputum culture. Stable COPD patients could not be included in the study because of the study was performed in the emergency department. This may have limited our study.

Statistical method

For the statistical analysis of the data obtained, the SPSS (Statistics Program for Social Scientists) 24, 0 program was used. When the power analysis of the cases was performed, sufficient cases were reached between healthy and patient cases. The Mann–Whitney U test was used to compare the data that did not fit the normal distribution among the groups, and Student's t test was used to compare the normally distributed data. p < 0.05 was considered statistically significant. A χ^2 test was used to compare frequency data between two groups. p < 0.05 was considered statistically significant.

The Kruskal–Wallis test was used to compare three groups (acute exacerbation of COPD, acute exacerbation of COPD and pneumonia, healthy subjects) because the presepsin levels did not conform to a normal distribution. *Post hoc* analysis was performed with the Bonferroni test. Correlation between presepsin and inflammatory markers was investigated. The Spearman test was used to examine correlations. Med Calc statistical program was used for the receiver operating characteristic (ROC) analysis.

Results

The study included 126 patients and 44 healthy individuals. While 51.6% (n = 65) of the patients were diagnosed with acute exacerbation of COPD + pneumonia. There were 26 female and 39 male patients with acute exacerbation of COPD. There were 27 female and 34 male patients with acute exacerbation of COPD + pneumonia. Of the 126 patients, 42.1% were female and 57.9% were male. The mean age of the patients with acute exacerbation of COPD (group 1) was 66.35 \pm 13.1 (min 31–max 95); the mean age of the patients with COPD acute exacerbation + pneumonia (Group 2) was 72.82 \pm 11.38 (min 40–max 91). The mean values of age and vital signs of all patients included in the study are shown in Table 1. A statistically significant difference was found between the ages of group 1 (COPD acute exacerbation) and group 2 (COPD acute exacerbation + pneumonia; p < 0.05; Table 1). There was a significant difference between groups for systolic blood pressure, diastolic blood pressure, pulse, fever, respiratory rate or oxygen saturation (p > 0.05; Table 1). Significant correlations were found between presepsin and systolic BP (p = 0.024; r: -0.201), diastolic BP (p = 0.035; r: -0.182), pulse (p = 0.008; r: 0.235), fever (p = 0.027; r: 0.197), oxygen saturation (p = 0.001; r: -0.282). There was no correlation between presepsin and respiratory rate.

Hypertension was the most common cause of co-morbid diseases, with a prevalence of 42.9% in the patients included in the study, followed by diabetes with a prevalence of 20.6% (Table 2). There was no statistically significant difference in co-morbidity between the groups (p > 0.05).

61 of the patients included in the study were active smokers. There was no significant difference between presepsin levels of smokers (5.33 ± 5.3 ; min: 0.1-max: 27.8) and nonsmokers (6.16 ± 5.9 ; min: 0.1-max: 27.8; p = 0.4). No significant difference was found between the two patient groups in terms of smoking, presence of oxygen condensers, living in a home for the aged or history of intubation. In addition, 51.6% of the total patients were using oxygen condensers at home. The rate of smoking among all patients was 48.4%. There was no statistically significant difference in corticosteroid and $\beta 2$ agonist drug use between the two groups (Table 2). When the symptoms of the cases were examined, cough was found in 65.1%, followed by sputum in 63.5% of patients. There was a significant difference only in cracles in terms of physical examination findings (p < 0.05; Table 2).

Chest x-ray and CT were performed in 53 patients. Chest x-ray was performed in 73 patients. In the COPD + pneumonia group, 67.2% (n = 41) of the patients had x-ray + tomography and 32% (n = 20) had chest x-ray.

There was a significant difference between the two patient groups in terms of white blood cells (WBC), urea, creatine and PCO₂ values (p < 0.05). There was no statistically significant difference between the other blood parameters (p > 0.05; Table 1). Significant correlations were found between presepsin and WBC (p = 0.015; r:

Vital signs and laboratory values	Acute exacerbation of COPD (n = 65)	Acute exacerbation of COPD + pneumonia (n = 61)	p-value
Age (years)	66.3 ± 13.1 (31–95)	72.8 ± 11.4 (40–91)	0.003*
Systolic blood pressure (mm/Hg)	134.1 ± 27.1 (90–220)	122.4 \pm 21.5 (70–170)	0.44
Diastolic blood pressure (mm/Hg)	76.83 ± 13.84 (50–113)	71.18 ± 13.30 (40–115)	0.20
Pulse rate/minute	93.64 ± 18.84 (18–130)	100 ± 19.60 (69–150)	0.12
Body temperature	36.82 ± 0.528 (36–38.5)	37.1 ± 0.94 (35.6–95)	0.214
Respiratory rate/minute	21.58 ± 4.05 (12–32)	20.57 ± 3.60 (12–30)	0.252
Pulse oxygen saturation	86.44 ± 11.85 (52–100)	84.3 ± 10.88 (54–99)	0.130
White blood cell	10,173 \pm 3502 (4500–20,000)	13,555 \pm 6875 (4800–41,200)	0.001*
Hemoglobin	13.85 ± 2.168 (7.5–18.20)	13.31 ± 2.28 (7.2–17.9)	0.122
Hematocrit	42.50 ± 8.53 (4.80–58.80)	41.68 ± 7.39 (23.10–59.80)	0.339
Platelets	235,507 \pm 81,641 (68,000–518,000)	260,278 ± 128,783 (98 000–676,000)	0.712
Jrea	47.69 ± 28.39 (13–155)	61.32 ± 36.13 (16–208)	0.014*
Creatinine	1.06 \pm 0.336 (0.61–1.94)	1.33 ± 0.889 (0.52–6.97)	0.029*
AST	104.5 ± 465.53 (9–3388)	40.73 ± 51.50 (13–340)	0.213
ALT	103.2 \pm 479.65 (6–3520)	36.86 ± 65.66 (5–336)	0.498
Glucose	134.33 \pm 53.24 (52–360)	155.09 \pm 74.56 (82–495)	0.096
Sodium	136.47 ± 3.67 (125–142)	135.83 ± 4.52 (124–145)	0.272
Potassium	4.44 ± 0.571 (3.30–6.14)	$4.55 \pm 0.719 \text{ (3.066.03)}$	0.253
Н	$7.34 \pm 0.83 \ \textbf{(7.07-7.70)}$	7.34 ± 0.84 (7.17–7.51)	0.758
5pO2	64.80 ± 22.59 (15–97.9)	64.57 ± 22.68 (12.7–98.2)	0.973
PaO ₂	46.2 ± 22.16 (15.6–135)	43.95 ± 17.50 (1.90–91.30)	0.903
PCO ₂	58.76 ± 15.26 (29.20–96.80)	52.81 \pm 16.86 (22.80–96.20)	0.048*
FEV1 %	46.5 ± 21.1 (12.6–91.7)	51.6 ± 18.4 (22.2–95.3)	0.23
EV1/FVC %	47.01 ± 14.4 (53.5–69.4)	47.5 ± 15.4 (34.7–69.2)	0.8
GOLD stage	2.8 ± 0.9 (1–4)	2.5 ± 0.7 (1–4)	0.15 [†]
Number of exacerbation in the previous	3 ± 6 (0–29)	3 ± 7 (0–34)	0.28

Data was shown as mean \pm SD (min-max) unless otherwise stated. p-values were calculated by Mann-Whitney U test.

[†]Student's t test.

*Statistically significant p-values (<0.05).

COPD: Chronic obstructive pulmonary disease; FEV1: Forced expiratory volume in 1 s; FVC: Forced vital capacity; GOLD: Global initiative for chronic obstructive lung disease; PaO₂: Partial oxygen pressure; PCO₂: Partial carbon dioxide pressure; SD: Standard derivation; SpO₂: Oxygen saturation.

0.282), hemoglobin (p = 0.014; r: -0.219), hematocrit (p = 0.016; r: -0.215), urea (p = 0.000; r: 0.443), creatinine (p = 0.000; r: 0.438), AST (p = 0.000; r: 0.318), sodium (p = 0.001; r: -0.303), potassium (p = 0.005; r: 0.251). There was no correlation between presepsin and glucose, ALT, platelets and blood gas values.

There was no statistically significant difference in forced expiratory volume in 1 s (FEV1; p = 0.23), FEV1/forced vital capacity (FVC; p = 0.8), GOLD (p = 0.15) between the groups (Table 1). According to the GOLD staging, 8% of the patients were stage 1, 32% were stage 2, 44% were stage 3 and 16% were stage 4.

There was no significant correlation between the levels of presepsin and FEV1 (r = 0.248; p = 0.08), FEV1/FVC (r = 0.121; p = 0.40).

The mean number of exacerbation in the previous 12 months was 3 ± 6 (min 0-max 34). There was no significant difference in the number of exacerbation in the previous 12 months between the groups (p = 0.28; Table 1). The exacerbations of the patients included in the study were not added to these numbers.

The values of ESR, CRP and procalcitonin were compared statistically between groups. ESR, CRP and procalcitonin values were significantly higher in the COPD acute exacerbation + pneumonia group (p < 0.05; Table 3).

Presepsin levels were significantly higher in the COPD acute exacerbation + pneumonia group than in the COPD acute exacerbation group (p < 0.05; Table 3 & Figure 1). Presepsin levels of the three groups were compared statistically, and a statistically significant difference was determined (p < 0.05). A *post hoc* analysis Bonferroni test was used to determine by which groups the difference was caused. The level of presepsin in the

Characteristics	All patients (n = 126)	Acute exacerbation of COPD (n = 65)	Acute exacerbation of COPD + pneumonia (n = 61)	p-value
Co-morbidity				
Diabetes mellitus	26 (20.6)	14 (21.5)	12 (19.7)	0.80
Hypertension	54 (42.9)	24 (36.9)	30 (49.2)	0.16
Cardiovascular disease	37 (29.4)	19 (29.2)	18 (29.5)	0.97
Congestive heart failure	26 (20.6)	11 (16.9)	15 (24.6)	0.29
Cerebrovascular disease	3 (2.4)	2 (3.1)	1 (1.6)	0.6
Chronic renal disease	8 (6.3)	4 (6.2)	4 (6.6)	0.92
History				
Smoker	61 (48.4)	32 (49.2)	29 (47.5)	0.85
Use of oxygen condenser (supplemental oxygen)	65 (51.6)	33 (50.8)	32 (52.5)	0.85
Stay in the nursing home	15 (11.9)	4 (6.2)	11 (18.0)	0.40
History of intubation	2 (1.6)	1 (1.5)	1 (1.6)	0.96
Use of β2 agonist	92 (73)	50 (76.9)	42 (68.8)	0.29
Use of corticosteroid	78 (61.9)	42 (64.6)	36 (59)	0.47
Symptoms and physical examination	tion findings			
Increase of sputum production	80 (63.5)	39 (60.0)	41 (67.2)	0.4
Cough	82 (65.1)	42 (64.6)	40 (65.6)	0.91
Fever	41 (32.5)	20 (30.8)	21 (34.4)	0.66
ncrease of sputum purulence	31 (24.6)	16 (24.6)	15 (24.6)	0.99
Wheezing	65 (51.6)	44 (67.7)	21 (34.4)	0.91
Crackles	68 (54)	20 (30.8)	48 (78.6)	0.04*
Mortality	36 (28.6)	20(30.8)	16 (26.2)	0.57

*p-values statistically significant.

COPD: Chronic obstructive pulmonary disease.

Table 3. Comparison of inflammatory markers of patients.

Inflammatory markers	Acute exacerbation of COPD (n = 65)	Acute exacerbation of COPD + pneumonia (n = 61)	p-value
Presepsin (ng/ml)	$3.62 \pm 3.08 \text{ (0.118.8)}$	8.03 ± 6.69 (0.2–27.8)	<0.001
Procalcitonin (ng/ml)	1.09 ± 6.94 (0–56)	5.65 ± 16.37 (0–100)	<0.001
CRP (mg/l)	44.54 \pm 60.03 (0.48–299)	98.43 ± 82.99 (7.35–312)	<0.001
ESR (mm/h)	26.66 ± 19.0 (2–90)	36.03 ± 25.06 (120)	0.03
Data shown as mean + SD (min-max)	inless otherwise stated in-values were calculated by Mar	n–Whitney U test $n < 0.05$ statistically significant	

Data shown as mean \pm SD (min–max) unless otherwise stated. p-values were calculated by Mann–Whitney U test, p < 0.05 statistically significant. COPD: Chronic obstructive pulmonary disease; ESR: Erythrocyte sedimentation rate; SD: Standard derivation.

Table 4. Relations	hip of presepsi	n with mortality ar	nd other inflammatory markers.	
Presepsin		ESR	CRP	Procalcitonin
Presepsin	r	0.209	0.482	0.590
	р	0.02	<0.001	<0.001
Spearman correlation test, p ESR: Erythrocyte sedimentati		nificantly.		

group with acute exacerbation of COPD acute exacerbation + pneumonia was statistically significantly higher than the levels in both the healthy group and in the COPD acute exacerbation group (p < 0.001; Figure 1).

A significant positive correlation was found between the levels of presepsin and ESR, CRP and procalcitonin (Table 4).

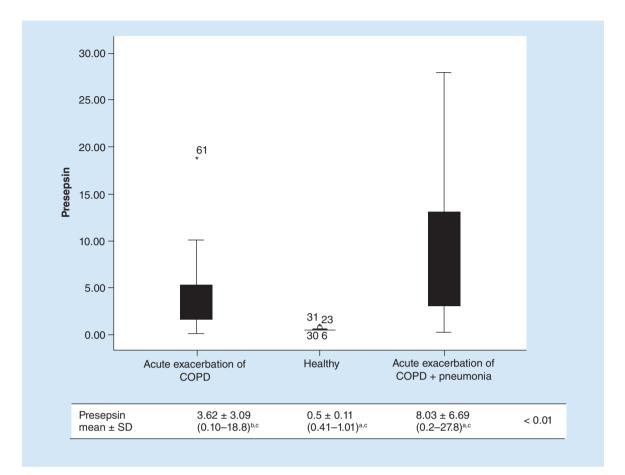


Figure 1. Comparison of presepsin levels between three groups. ^aIndicates the statistical difference to acute exacerbation of COPD. ^bIndicates the statistical difference to acute exacerbation of COPD + pneumonia, ^cIndicates the statistical difference to healthy individual. Kruskal Wallis Test, p < 0.05 statistical significant, post hoc Benforrini test was used.

COPD: Chronic obstructive pulmonary disease.

Table 5. Presepsin levels in patients with and without mortality.				
Groups	Patients with mortality	Patients without mortality	p-value	
All patients	$6.36 \pm 5.53 \text{ (0.118.4)}$	$5.52 \pm 5.63 \; \textbf{(0.1-27.8)}$	0.33	
AECOPD	3.98 \pm 2.77 (0.1–10)	$\textbf{4.47} \pm \textbf{3.23} \; \textbf{(0.1-18.8)}$	0.33	
AECOPD + Pneumonia	$9.33 \pm 6.69 \text{ (1.2-18.40)}$	7.57 ± 6.71 (0.2–27.8)	0.35	
		ulated by Mann–Whitney U test, $p < 0.05$ statistically significa rocyte sedimentation rate; SD: Standard derivation.	int.	

Receiver operating characteristic (ROC) analysis was performed for the ability of presepsin and other inflammatory markers to determine the possibility of pneumonia in COPD patients. The area under the curve (AUC) site was the highest in CRP, followed by procalcitonin, presepsin and ESR (Table 5 & Figure 2). The AUC area of the presepsin was 0.70, and the cut-off value was 7. When the cut-off value of 7 was used, the specificity of the presepsin was 92.31% and the sensitivity was 42.62% (Table 5).

The mortality rate was 28.6% (n = 36). The mortality rate in group 1 (acute exacerbation of COPD) was 30.8% (n = 20), while in group 2 (COPD acute exacerbation and pneumonia), the mortality rate was 26.2% (n = 16). There was no statistically significant difference between the two groups in terms of mortality (p > 0.05; Table 2). In addition, presepsin levels in patients with and without mortality were compared in all cases. There was no statistically significant difference in presepsin levels between the groups with and without mortality in

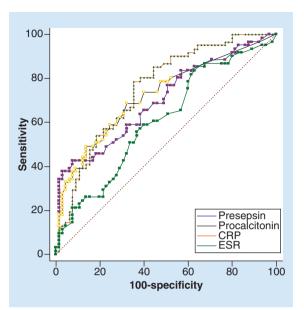


Figure 2. Comparison of presepsin, procalcitonin, CRP and erythrocyte sedimentation rate values for receiver operating characteristic curves in determining pneumonia.

ESR: Erythrocyte sedimentation rate.

Table 6. Receiver operating characteristic analysis of inflammatory biomarkers to determine the possibility of

prieumorna ir	i chi onic obsti	uctive pulmonary di	sease patients.			
Inflammatory Markers	AUC	Sensitivity	Specificity	Cut-off	SE	95% CI
Presepsin	0.70	42.62	92.31	7	0.0467	0.613-0.780
Procalcitonin	0.72	68.85	67.69	0.16	0.0447	0.636-0.799
CRP	0.75	78.69	64.62	25.7	0.0431	0.668-0.826
ESR	0.62	85.25	35.38	15	0.0505	0.518-0.695
ALIC: Area under the	curvo: ESR: Endbroch	to codimontation rate: SE: Star	dard Error			

AUC: Area under the curve; ESR: Erythrocyte sedimentation rate; SE: Standard Error.

Table 7. Comparison of inflammatory markers according to sputum culture results.			
Inflammatory markers	Culture negative (n = 37)	Culture positive (n = 15)	p-value
Presepsin (ng/ml)	8.21 \pm 6.30 (1.20–27.8)	8.14 \pm 8.91 (0.6–27.8)	0.57
Procalcitonin (ng/ml)	3.05 ± 6.14 (0–22)	11.41 \pm 33.22 (0–100)	0.71
CRP (mg/l)	116.66 \pm 100.01 (9.2–312)	107.3 \pm 82.63 (23.8–236)	0.96
ESR (mm/h)	35.00 ± 22.97 (3–78)	46.44 ± 31.43 (17–120)	0.40

the COPD acute exacerbation + pneumonia group. The same results were obtained in the acute exacerbation of COPD group (Table 4).

Sputum or tracheal aspirate samples were taken from 52 patients with COPD acute exacerbation + pneumonia, and the cultures were studied. In all, 32.7% (n = 17) of the patients had no growth in their cultures. Normal respiratory tract flora was found in 38.5% (n = 20) of the patients, and 28.8% (n = 15) of the patient cultures showed growth of pathogen microorganisms. The groups with positive cultures and negative cultures + normal respiratory flora was compared with each other. No statistically significant difference was found between the two groups in presepsin (p = 0.57), procalcitonin (p = 0.71), CRP (p = 0.96) or ESR (p = 0.40; Table 6). In the 20 blood cultures performed, no growth was observed.

The distribution of pathogenic microorganisms was as follows: Gram-negative microorganisms were obtained in eight patients (*Pseudomonas aeruginosa, Stenotrophomonas maltophilia, Acinotobacter haemolyticus, Klebsiella Pneumoniae, Gram-negative laktobasil, Hemofilus influenza*), Gram-positive microorganisms in six patients (*Streptococcus pneumonia, Staphylococcus aureus*) and *Candida albicans* in one case. No statistically significant difference was found

Inflammatory markers	Gram-negative bacteria (n = 8)	Gram-positive bacteria (n = 6)	p-value
Presepsin (ng/ml)	7.42 ± 6.67 (0.6–17.7)	9.32 ± 9.67 (1.5–27.8)	0.67*
Procalcitonin (ng/ml)	7.98 ± 21.82 (0–62)	17.31 \pm 40.52 (0–100)	0.65
CRP (mg/l)	122.25 ± 90.95 (23.8–285)	167.19 \pm 127.95 (7.87–312)	0.46*
ESR (mm/h)	48 ± 33.4 (17–120)	46.7 ± 32.8 (2–82)	0.94*

between presepsin levels of patients with pneumonia due to Gram-positive $(9.32 \pm 9.67; \text{min: } 1.5-\text{max: } 27.8)$ and Gram-negative $(7.42 \pm 6.67; \text{min: } 0.6-\text{max: } 17.7)$ bacteria (p = 0.67; Table 6).

Discussion

In our study, we wanted to investigate the role of presepsin as a new infectious mediator in patients with acute exacerbation of COPD. We evaluated the relationship of presepsin with procalcitonin and acute phase reactants (ESR and CRP), which are accepted markers used for antibiotic usage guidelines in bacterial infections.

In many studies reported that CRP and procalcitonin levels were significantly higher in the COPD exacerbation with pneumonia group compared with those of the COPD exacerbation group [7,8]. Taghizadieh *et al.* compared CRP and procalcitonin biomarkers in patients with COPD exacerbation and community-acquired pneumonia. They found a statistically significant difference in procalcitonin in the community-acquired pneumonia group compared with the COPD group but reported no significant difference in CRP values [9]. In a study by Klouche *et al.*, patients with acute respiratory failure, patients with sepsis due to pneumonia and noninfectious patients were compared. CRP and procalcitonin (cut-off: 0.5 ng/ml) were significantly higher in patients with sepsis due to pneumonia [16]. Since CRP and procalcitonin infection markers were accepted, most of the studies found these markers to be higher in patients with COPD + pneumonia. Both parameters are used to determine the presence of infection in patients presenting with acute exacerbation of COPD. In our study, procalcitonin and CRP levels in the COPD acute exacerbation + pneumonia group were found to be statistically significant. However, there are other studies in the literature with differing findings. Kawamatawong *et al.* did not find a significant difference in procalcitonin and CRP levels between bacterial infections and other etiologies (viral and noninfectious) in patients with acute exacerbations of COPD. They stated that the small number of patients in their study may have affected this result [10].

It has been shown in many studies that ESR, an inflammatory marker, is increased in COPD acute exacerbations and community-acquired pneumonia [17–20]. However, we did not find any study comparing ESR levels in patients with pneumonia in COPD acute exacerbation and in patients with acute exacerbation of COPD. In a study performed in COPD patients with lower respiratory tract infection, Van Beurden *et al.* reported that ESR increased and that this increase decreased with treatment [19]. In our study, ESR was found to be statistically higher in the group of patients with COPD acute exacerbation and pneumonia compared with the group without pneumonia. Acute exacerbation of COPD with increased inflammation is associated with pneumonia, leading to a further increase in ESR.

In this study, compared with healthy people and patients with acute exacerbation of COPD, presepsin was significantly higher in the acute exacerbation of COPD group. In the literature, there was no study showing the level of presepsin in patients with acute exacerbation of COPD. However, recent studies have shown that the level of presepsin increases during inflammation and infection [16,21]. High levels of presepsin in COPD may be attributed to acute exacerbation characterized by increased inflammatory mediators and airway inflammation. This may be the reason why presepsin levels increase above the cut-off value (7 ng/ml) in some of the patients with acute exacerbation of COPD. Since we performed the study in the emergency department, we could not include stable COPD patients. The lack of stable COPD patients has limited our study. More studies are needed to fully evaluate the potential role of presepsin in the diagnosis of acute exacerbation of COPD.

We compared the value of presepsin in acute exacerbation of COPD and acute exacerbation of COPD + pneumonia groups. The value of serum presepsin was significantly higher in patients with acute exacerbation of COPD with pneumonia. No similar study was found in the literature. However, Klouche *et al.* compared the levels of presepsin to differentiate patients with sepsis and noninfectious respiratory failure [16]. Presepsin (cut-off value: 588 pg/ml) was found to be significantly higher in patients with sepsis due to pneumonia [16]. There are also reports in the literature that presepsin can be used in the diagnosis and prognosis of patients with pneumonia [16,22,23]. In one study, Liu *et al.* investigated the role of presepsin in determining the severity of community-acquired pneumonia; they found that presepsin levels in patients with severe pneumonia were significantly higher than those in the other pneumonia cases [22]. In a study by Qi *et al.*, the levels of blood presepsin between community-acquired bacterial pneumonia and active tuberculosis patients were compared. In this study, the presepsin level of the pneumonia group was found to be significantly higher than that of the active tuberculosis group [23]. Proteolysis of sCD14 released into the circulation by cathepsin leads to presepsin formation. Thus, presepsin levels in the blood can indicate inflammation and infection. Presepsin can be detected in the blood as long as the infection continues. Presepsin, which may increase in acute exacerbation of COPD due to inflammation, was significantly increased with the addition of pneumonia.

It has been shown in several studies that concentrations of presepsin increase with age and renal impairment [24,25]. In the present study, there was a significant but weak correlation between presepsin and the ages of the cases. In additionally, the statistically significant difference was found between the ages of group 1 (COPD acute exacerbation) and group 2 (COPD acute exacerbation + pneumonia). The effect of age on the difference between presepsin levels of these two groups should also be considered. Age should be considered in the interpretation of presepsin levels and threshold values should be established accordingly.

Huerta *et al.* have reported that corticosteroid use affects patients' inflammatory markers [7]. In our study, no significant difference was found between the two groups in terms of corticosteroid use. Therefore, we think that steroid has no role in the levels of presepsin and other inflammatory markers.

Titova *et al.* examined the area under the ROC curve of procalcitonin (0.67) and CRP (0.73) to detect pneumonia [8]. In our study, the highest AUC value was found in CRP (AUC = 0.75), in accordance with the literature. In our study, the values of procalcitonin, ESR and presepsin were found to be 0.72, 0.61 and 0.70, respectively. In the study by Klauche *et al.*, presepsin and procalcitonin tests were performed for the diagnosis of pneumonia in patients admitted to the intensive care unit with acute respiratory failure. As a result of the analysis, the AUC value of presepsin was 0.75, the sensitivity was 81% and the specificity was 80%. The AUC value of procalcitonin AUC values were higher than those in our study. We attributed this difference to the fact that in the Klauche *et al.* study, all the patients with pneumonia were patients with sepsis. In recent studies, levels of presepsin were found significantly higher in septic than in nonseptic patients or those with systemic inflammatory response syndrome. Moreover, a specific increase was reported in the early stage of sepsis that also well correlated with severity. Especially in severe sepsis, impaired renal function and bleeding parameters increase presepsin levels more [16,26,27]. Since sepsis and septic shock patients were not included in this study, the levels of inflammatory markers may not be increased further.

In this study; FEV1, FEV1/FVC and GOLD values of the patients in the two groups were similar. Additionally, there was no relationship between the severity of COPD and presepsin levels. Therefore, we do not think that the degree of COPD affects the level of presepsin in our study.

In the study by Klauche *et al.*, patients with sepsis who had severe pneumonia had significantly higher values of presepsin and procalcitonin than those living with sepsis alone. There was no statistically significant difference in CRP values [16]. In our study, no significant difference was found in terms of mortality in both groups. We think that the result of the study by Klauche *et al.* was because all patients had severe pneumonia with sepsis. We did not include sepsis patients in the study.

Lacoma *et al.* reported that 39% of the patients with COPD exacerbations had normal flora, 35% had pathogen microorganisms and 24% had no growth in culture [6]. They found no significant difference in procalcitonin and CRP levels when compared with the normal flora + culture-negative and culture-positive groups. They did not find any significant difference between the etiologies of infection for either marker [6]. Titova *et al.* reported that in the sputum cultures of COPD patients with acute exacerbation and patients with COPD acute exacerbation + pneumonia, 27% had normal flora microorganisms and 30% had pathogenic microorganisms. They also showed that 42% of the patients were infected with viruses. Procalcitonin levels and these pathogens have not been reported to be correlated statistically. There was no growth in the blood cultures of the patients [8]. Our study was consistent with the above studies; 32.7% of the patients had no growth in culture. Normal respiratory tract flora was found in 38.5% of the patients, and 28.8% of the patients developed pathogen microorganisms. In the culture, the pathogen-producing group and the nonpathogen groups were compared; no statistically significant difference

was found for presepsin, procalcitonin, CRP or ESR levels. Endo *et al.* reported that there was no significant difference between presepsin levels in infections due to Gram-positive and Gram-negative pathogens [28]. Our results were consistent with this study. These findings indicate that infection and colonization cannot be differentiated according to culture results. In addition, none of the inflammatory markers used seems to be useful in determining the etiology of infection.

Conclusion

ESR, CRP, procalcitonin and presepsin levels were significantly higher in patients with COPD acute exacerbation + pneumonia compared with those of patients with isolated COPD acute exacerbation. Presepsin was significantly correlated with the other three inflammatory markers.

As a result, presepsin can be used to diagnose pneumonia in patients with acute exacerbation of COPD admitted to the emergency department.

Summary points

- The level of presepsin in the group with acute exacerbation of chronic obstructive pulmonary disease (COPD)
 + pneumonia was statistically significantly higher than the levels in both the healthy group and in the COPD
 acute exacerbation group. Presepsin levels were significantly higher in the COPD acute
 exacerbation + pneumonia group than in the COPD acute exacerbation group.
- Erythrocyte sedimentation rate (ESR), CRP and procalcitonin values were significantly higher in the COPD acute exacerbation + pneumonia group.
- A significant positive correlation was found between the levels of presepsin and ESR, CRP and procalcitonin.
- ROC analysis was performed for the ability of presepsin and other inflammatory markers to determine the possibility of pneumonia in COPD patients. The AUC area was the highest in CRP, followed by procalcitonin, presepsin and ESR.
- The groups with positive cultures and negative cultures + normal respiratory flora were compared with each other. No statistically significant difference was found between the two groups in inflammatory markers.

Author contributions

The authors contributed equally to this study. Constructing an idea or hypothesis for research: S Özkan, A Halıcı. Planning methodology: S Özkan, A Halıcı. Data collection: A Halıcı, İ Hür, K Abatay, F Halıcı. Biochemical analysis: E Çetin. Statistical analysis: S Özkan. Writing: A Halıcı, S Özkan.

Financial & competing interests disclosure

The authors have no relevant affiliations or financial involvement with any organization or entity with a financial interest in or financial conflict with the subject matter or materials discussed in the manuscript. This includes employment, consultancies, honoraria, stock ownership or options, expert testimony, grants or patents received or pending, or royalties.

No writing assistance was utilized in the production of this manuscript.

Ethical conduct of research

This observational prospective study was carried out in line with research regulations, including the approval of the Ethics Committee of our institute dated 19/03/2018 and numbered 47/10 and according to the principles of the 'World Medical Association Helsinki Declaration.'

References

Papers of special note have been highlighted as: • of interest; •• of considerable interest

- 1. Celli BR, Vestbo J. The EXACT-pro: measuring exacerbations of COPD. Am. J. Respir. Crit. Care Med. 183(3), 287-288 (2011).
- 2. Vogelmeier CF, Criner GJ, Martinez FJ *et al.* Global strategy for the diagnosis, management, and prevention of chronic obstructive lung disease 2017 report. GOLD executive summary. *Am. J. Respir. Crit. Care Med.* 195(5), 557–582 (2017).
- 3. Wedzicha JA, Seemungal TA. COPD exacerbations: defining their cause and prevention. Lancet 370(9589), 786–796 (2007).
- 4. Sapey E, Stockley RA. COPD exacerbations. 2: aetiology. Thorax 61(3), 250-258 (2006).
- Restrepo MI, Mortensen EM, Pugh JA et al. COPD is associated with increased mortality in patients with community-acquired pneumonia. Eur. Respir. J. 28, 346–351 (2006).

- Lacoma A, Prat C, Andreo F et al. Value of procalcitonin, C-reactive protein, and neopterin in exacerbations of chronic obstructive pulmonary disease. Int. J. Chron. Obstruct. Pulmon. Dis. 6, 157–169 (2011).
- 7. Huerta A, Crisafulli E, Menendez R *et al.* Pneumonic and nonpneumonic exacerbations of COPD: inflammatory response and clinical characteristics. *Chest* 144(4), 1134–1142 (2013).
- Compared inflammatory markers between pneumonia and nonpneumonic cause in chronic obstructive pulmonary disease (COPD) acute exacerbations.
- Titova E, Christensen A, Henriksen AH, Steinshamn S, Asberg A. Comparison of procalcitonin, C-reactive protein, white blood cell count and clinical status in diagnosing pneumonia in patients hospitalized with acute exacerbations of COPD: a prospective observational study. *Chron. Respir. Dis.* 16, 1479972318769762 (2019).
- •• Similar to our study, the role of procalcitonin in the diagnosis of pneumonia associated with acute exacerbation of COPD was investigated.
- 9. Taghizadieh A, Soleimanpour H, Rahmani F, Nia KS, Khodaverdizadeh H. Comparison of serum levels of procalcitonin and C-reactive protein in patients with community acquired pneumonia and COPD exacerbation. *Iran Red. Crescent. Med. J.* 18(12), e25811 (2016).
- 10. Kawamatawong T, Apiwattanaporn A, Siricharoonwong W. Serum inflammatory biomarkers and clinical outcomes of COPD exacerbation caused by different pathogens. *Int. J. Chron. Obstruct. Pulmon. Dis.* 12, 1625–1630 (2017).
- 11. Su YJ. The value of C-reactive protein in emergency medicine. J. Acute Dis. 3(1), 1-5 (2014).
- 12. Shirakawa K, Naitou K, Hirose J, Takahashi T, Furusako S. Presepsin (sCD14-ST): development and evaluation of one-step ELISA with a new standard that is similar to the form of presepsin in septic patients. *Clin. Chem. Lab. Med.* 49(5), 937–939 (2011).
- Mussap M, Puxeddu E, Burrai P et al. Soluble CD14 subtype (sCD14-ST) presepsin in critically ill preterm newborns: preliminary reference ranges. J. Matern. Fetal Neonatal. Med. 25(Suppl. 5), 51–53 (2012).
- 14. Liu B, Chen YX, Yin Q, Zhao YZ, Li CS. Diagnostic value and prognostic evaluation of presepsin for sepsis in an emergency department. *Crit. Care.* 17(5), R244 (2013).
- A study on the importance of presepsin in infection and sepsis in the emergency department.
- 15. Romualdo LG, Torrella PE, Gonzales MV *et al.* Diagnostic accuracy of presepsin (soluble CD14 subtype) for prediction of bacteremia in patients with systemic inflammatory response syndrome in the emergency department. *Clin. Biochem.* 47(7–8), 505–508 (2014).
- 16. Klouche K, Cristol JP, Devin J et al. Diagnostic and prognostic value of soluble CD14 subtype (presepsin) for sepsis and community-acquired pneumonia in ICU patients. Ann. Intensive Care. 6(1), 59 (2016).
- Reports that presepsin can be used to diagnose severe community-acquired pneumonia, sepsis and severe sepsis in patients admitted to the intensive care unit due to acute respiratory failure.
- 17. Taylan M, Demir M, Selimoglu Sen H *et al.* Alterations of the neutrophil-lymphocyte ratio during the period of stable and acute exacerbation of chronic obstructive pulmonary disease patients. *Clin. Respir. J.* 11(3), 311–317 (2017).
- In E, Kuluöztürk M, Öner Ö, Deveci F. The importance of neutrophil-to-lymphocyte ratio in chronic obstructive pulmonary disease. *Turk. Thorac. J.* 17(2), 41–46 (2016).
- 19. van Beurden WJ, Smeen FW, Harff GA, Dekhuijzen PN. Markers of inflammation and oxidative stress during lower respiratory tract infections in COPD patients. *Monaldi. Arch. Chest Dis.* 59(4), 273–280 (2003).
- Hopstaken RM, Muris JW, Knottnerus JA, Kester AD, Rinkens PE, Dinant GJ. Contributions of symptoms, signs, erythrocyte sedimentation rate, and C-reactive protein to a diagnosis of pneumonia in acute lower respiratory tract infection. *Br. J. Gen. Pract.* 53(490), 358–364 (2003).
- 21. Anas A, van der Poll T, de Vos AF. Role of CD14 in lung inflammation and infection. Crit. Care. 14(2), 209 (2010).
- 22. Liu B, Yin Q, Chen YX, Zhao YZ, Li CS. Role of presepsin (sCD14-ST) and the CURB65 scoring system in predicting severity and outcome of community-acquired pneumonia in an emergency department. *Respir. Med.* 108(8), 1204–1213 (2014).
- Reports that presepsin can be used to determine the severity of community-acquired pneumonia in the emergency department.
- Qi ZJ, Yu H, Zhang J, Li CS. Presepsin as a novel diagnostic biomarker for differentiating active tuberculosis from bacterial community acquired pneumonia. *Clin. Chim. Acta* 478, 152–156 (2018).
- 24. Chenevier-Gobeaux C, Trabattoni E, Roelens M, Borderie D, Claessens YE. Presepsin (sCD14-ST) in emergency department: the need for adapted threshold values. *Clin. Chim. Acta* 1(427), 34–36 (2014).
- 25. Tong X, Cao Y, Yu M, Han C. Presepsin as a diagnostic marker for sepsis: evidence from a bivariate meta-analysis. *Ther. Clin. Risk Manag.* 2(11), 1027–1033 (2015).
- 26. Yaegashi Y, Shirakawa K, Sato N *et al.* Evaluation of a newly identified soluble CD14 subtype as a marker for sepsis. *J. Infect. Chemother.* 11(5), 234–238 (2005).
- 27. Behnes M, Bertsch T, Lepiorz D *et al.* Diagnostic and prognostic utility of soluble CD14 subtype (presepsin) for severe sepsis and septic shock during the first week of intensive care treatment. *Crit. Care.* 218(5), 507 (2014).
- 28. Endo S, Suzuki Y, Takahashi G *et al.* Usefulness of presepsin in the diagnosis of sepsis in a multicenter prospective study. *J. Infect. Chemother.* 18(6), 891–897 (2012).